

## Comparative Morphological Studies on NiO, CoO and Fe<sub>2</sub>O<sub>3</sub> Nanoparticles

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### Abstract

Stable oxides of Ni, Fe and Co nanoparticles are synthesized by chemical route using leaf extraction. Structural, biological morphological and magnetic properties were investigated. X ray pattern shows a polycrystalline structure of samples. Diffraction planes of (200), (122) and (200)-oriented crystal structures were revealed by X-ray pattern for NiO, Fe<sub>2</sub>O<sub>3</sub> and CoO respectively. Hexagonal, orthorhombic and cubic crystal structures have been revealed respectively for NiO, Fe<sub>2</sub>O<sub>3</sub> and CoO nanoparticles. Microscopic structure has been detected by using of Scanning Electron Microscopy. Ni, Fe and Co is present at 71%, 78.8%, 79% as indicated by EDS analysis respectively. Mass magnetization values of 51, 61 and 54 emu/g are recorded for NiO, Fe<sub>2</sub>O<sub>3</sub> and CoO nanoparticles.

**Keywords:** Ni; Fe; Co; Metal oxides; High transparency; Magnetic properties

### Introduction

The aim of this study was to find out comparative studies of metal oxides of Ni, Fe and Co mediated oxidative stress synthesized NPs. This study suggested that bare CoO NPs are a toxic for primary human immune cells that deals directly with human health. Surface modification or surface functionalization may open the gateway for further use of CoO NPs in different industrial use or in biomedical sciences [1,2]. The highest values of coercive field and saturation magnetization were obtained for the sample at  $x^{1/4}$ 1.0, while the lowest values were obtained in the sample with the highest Co excess ( $x^{1/2}$ 2.5). The results indicated that the used synthesis route was suitable for the synthesis of cobalt ferrite with moderate saturation magnetization and high coercive field values [3-5]. In addition, the de-amination is contributed by the nucleophilic attack of water molecule to the carbon atom of aromatic ring bonded to NH<sub>2</sub> group. After the de-amination reaction, the intermediate product is phenol. The following de-hydroxyl reaction by FeO domain proceeds immediately. The final product is benzene that is observed in all FIT involved catalytic routes [4]. Nanostructured iron oxides have been widely used in catalysis, adsorption, sensor, biomedicine, and so on. In this study, iron oxide nanoparticles containing nanosized cavities were prepared by hydrothermal treatment of a colloidal dispersion of iron hydroxide. Moreover, the iron oxide nanoparticles calcined 7-15 nm and 5-12 nm-diameter cavities, respectively. These peculiar structures might be conducive for some applications such as adsorption involving large molecules [5]. Pure NiO, CoO and Fe<sub>2</sub>O<sub>3</sub> exhibits anti ferromagnetic nature, which transforms into ferromagnetic one with moderate moment at room temperature with decreasing crystallite size. On the other hand, annealing of as-milled powders showed a large reduction in magnetic moment and the rate of decrease of moment strongly depends on the milling conditions. The observed properties are discussed on the basis of crystallite size variation, defect density, oxidation/reduction of Ni, Co and Fe and interaction between un- compensated surfaces and particle core with lattice expansion [6,7].

### Experimental Section

Materials used for the synthesis of Nickel oxide nanoparticles are Nickel Chloride salt (NiCl<sub>2</sub>), cobalt chloride CoCl<sub>2</sub> and ferrous chloride

FeCl<sub>2</sub> the salt was purchased from Merck, were the main sources of producing synthesized NiO, FeO and CoO, these three salt solutions were placed separately in three different beakers. These salt solutions were prepared in 1M ratio, with these individual beakers, 1mM NaOH was added separately 1  $\mu$ L of coreopsis leaf broth extract with, 30  $\mu$ L of milli Q water and then boiling the mixture before finally decanting it, freshly prepared double distilled water was used throughout the experimental work, the mixture was heated to 250°C for 20 minutes followed by filtering. Maintain this temperature for the entire reaction. Broth stored in refrigerator at 4°C. Within an hour (50 minutes) cherry red solution was obtained. Different plant extracts require different times to complete the reaction. The extract was added to  $3.5 \times 10^{-4}$  M Nickel chloride aqueous solution, obtaining different samples with different pH (the volumetric ratio mixture extract/Nickel chloride salt control sample containing only Nickel chloride salt. We have synthesized the Nickel oxide, cobalt oxide and ferrous oxide nanoparticles using this plant. The leaves were collected and the leaf broth was prepared by taking 5 g of thoroughly washed and finely cut leaves and 15  $\mu$ L of milli Q water. Then the leaves were nicely crushed in mortar pestle, transferred the nicely crushed leaves in to centrifuge tube and centrifuged at 10000-11000 rpm for 10 min at 4 to 5°C. 1  $\mu$ L of nitric acid was mixed with the 1  $\mu$ L of biological extract and added into 30  $\mu$ L of boiling milli Q water. Within a few minutes; the solution developed a distinct characteristic colour (Green) and later varied the volume of concentration (50  $\mu$ L and 100  $\mu$ L), like that the experiment is repeated for different pH (7, 9) by keeping the volume leaf extract (25  $\mu$ L) constant with constant stirring till the reaction got over (20 min) and studied the absorption of samples by using spectrophotometer.

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## Results and Discussion

### Structural characterization

The XRD pattern of NiO, CoO and Fe<sub>2</sub>O<sub>3</sub> were shown in Figure 1. All the observed diffraction peaks are indexed within face centred cubic (FCC) structure of NiO phase, in agreement with JCPDS card No. 451027 (Hexagonal) and FCC structure of CoO phase in agreement with JCPDS card-711178 (Cubic) and third nanoparticle Fe<sub>2</sub>O<sub>3</sub> phase in agreement with JCPDS-897047 Orthorhombic. It can be noticed

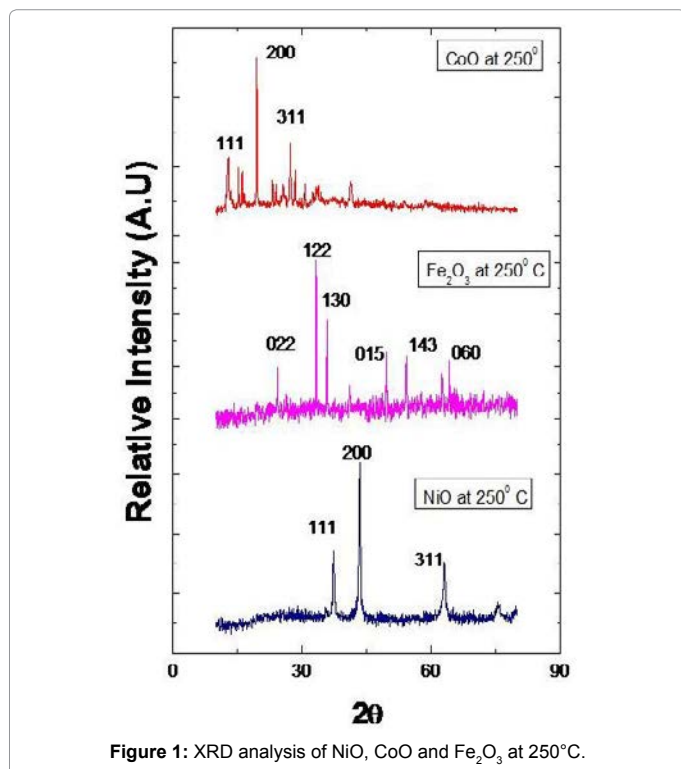


Figure 1: XRD analysis of NiO, CoO and Fe<sub>2</sub>O<sub>3</sub> at 250°C.

that the peak are broad, indicating the formation of nanocrystalline NiO, CoO and Fe<sub>2</sub>O<sub>3</sub> phase. In addition, with annealing temperature of 250°C, the intensity of peaks increases thereby enhanced crystallinity (Table 1). The crystallite size (D), dislocation density (δ), strain (ε), and lattice parameter (a), were calculated using the following formula:

$$D = \frac{0.9 \lambda}{\beta \cos \theta} \quad (1)$$

Where λ is the wavelength of the x-ray source, β is the full-width at half-maximum (FWHM in radian and θ is the Bragg's angle;

The dislocation parameter is then calculated;

$$\delta = \frac{1}{D^2} \quad (2)$$

The strain is then determined by the following relation,

$$\epsilon = \frac{\Delta d}{d_o} = \left( \frac{d_{\text{exp}} - d_{\text{hkl}}}{d_{\text{hkl}}} \right) \quad (3)$$

$$a = \frac{\Delta d}{d_o} \quad (4)$$

Where  $d_{\text{exp}}$  and  $d_{\text{hkl}}$  represent the experimental and for standard values of interplanar distance respectively calculated values are reported in Table 2. It can be noticed that the value of the crystallite size increases slightly setting in the range 29.82, 26.103 and 31.289 nm, due to grain growth. According to formula (2), the smaller value of the crystallite size, the larger the value of the dislocation density (inversely proportional), which indicates a better crystalline quality. From Table 1, it can be seen that the prepared films exhibit a compressive strain (all values are positive). In addition, when annealing temperature increases, the strain value increases, which is may be associated with the formation of oxygen vacancies [8].

### Scanning electron microscopy with EDAX

Morphological studies of microstructure analysis deals with agglomeration of the particles due to biosynthesis of the leaf extract.

Sample	Lattice constant Calculated (Å)			Crystal size size D= $\frac{0.94\lambda}{\beta \cos \theta}$ (nm)	Strain $\frac{\Delta d}{d_o} = \left( \frac{d_{\text{exp}} - d_{\text{hkl}}}{d_{\text{hkl}}} \right) \times 10^{-3}$ lines/m <sup>2</sup>	JCPDS
NiO	a=2.701	C=4.412		29.823	1.269	JCPDS -451027 Hexagonal $\frac{1}{d^2} = \frac{4}{3} \left[ \frac{h^2 + hk + k^2}{a^2} \right] + \frac{l^2}{c^2}$ Volume V= $\left( \frac{\sqrt{3}a^2C}{2} \right)$ Strain= $\beta \cos \theta / 4$
CoO	a=4.527			26.103	2.912	JCPDS-711178 a=4.263 Å <sup>0</sup> Cubic $d_{\text{hkl}} = \frac{a}{\sqrt{h^2 + k^2 + l^2}}$
Fe <sub>2</sub> O <sub>3</sub>	a=5.011	b=8.62	C=9.314	31.289	2.011	JCPDS -897047 Orthorhombic a=5.095 b=8.789 c=9.437 $\frac{1}{d^2} = \frac{h^2}{a^2} + \frac{k^2}{b^2} + \frac{l^2}{c^2}$ V= abc

Table 1: Structural analysis of NiO, CoO and Fe<sub>2</sub>O<sub>3</sub> Nanoparticles annealed @250°C.

Sample	Lattice constants Calculated (Å)			Crystal size size D=(nm)	Strain (x 10 <sup>-3</sup> lines/m <sup>2</sup> )
NiO	a=2.701		c=4.412	29.823	1.269
CoO	a=4.527			26.103	2.912
Fe <sub>2</sub> O <sub>3</sub>	a=5.011	b=8.62	c=9.314	31.289	2.011

Table 2: Structural and optical analysis of NiO, CoO and Fe<sub>2</sub>O<sub>3</sub> Nanoparticle at 250°C.

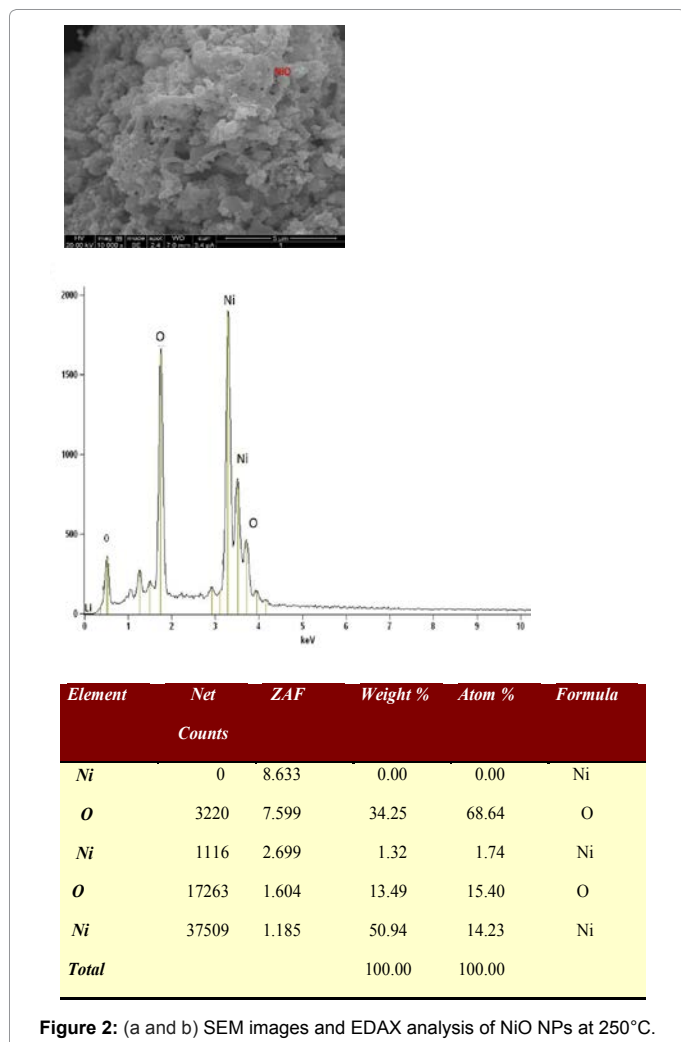


Figure 2: (a and b) SEM images and EDAX analysis of NiO NPs at 250°C.

Coreopsis leaf extract was added in the equal ratio of these three mother solutions of the particles. The as-prepared NPs were annealed at constant temperature of 250°C. The particles distributions of these NPs were almost constant shown in Figures 2a, 2b, 3a, 3b, 4a and 4b respectively. Scanning Electron Microscopy for creating nanofibers of these oxides materials and its elemental analysis confirmed the metal oxides and its presence in its condition (Figures 2-4).

### Transmission electron microscopy of NiO, FeO and CoO

Transmission Electron Microscopy confirmed the size distribution of metal oxides NPs which were below 100 nm, shown in Figure 5a-5c. The diffraction planes of x-ray diffractometer were well matched with SAED pattern. This efficient tool exhibits the real morphology of single particles and its agglomerations of the outcomes of the product. Slight dark batches present in these three particles distributions verify the biosynthesized products.

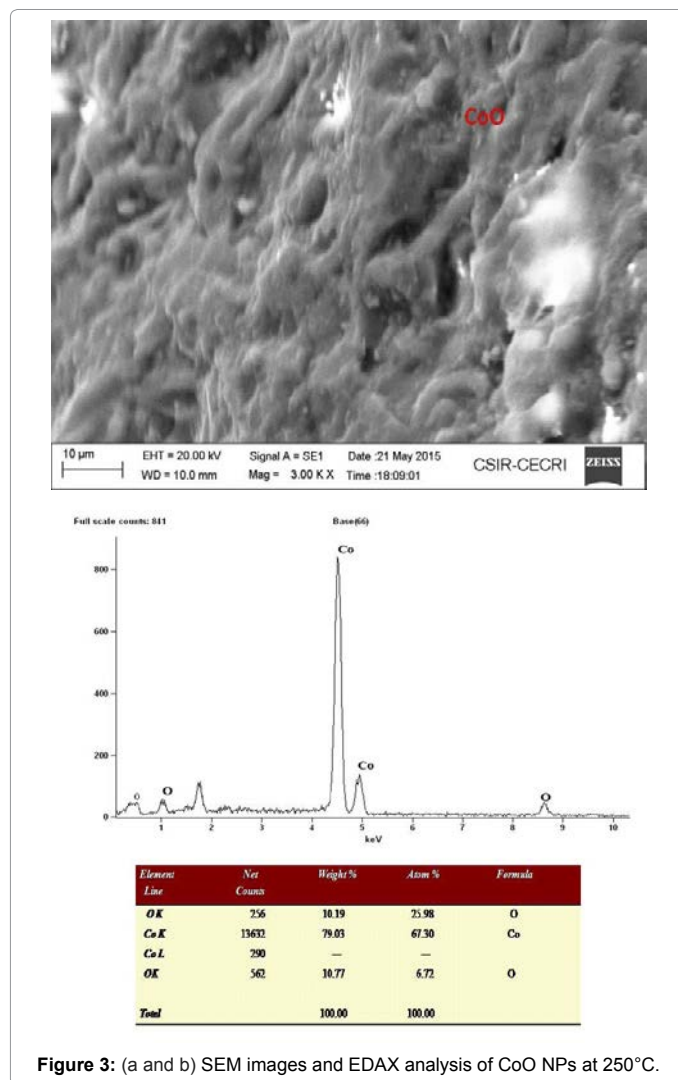


Figure 3: (a and b) SEM images and EDAX analysis of CoO NPs at 250°C.

### FT-IR analysis

Figure 6 illustrates the complementary information in the presence of NiO, CoO and Fe<sub>2</sub>O<sub>3</sub>, FT-IR analysis is observed on NiO, CoO and Fe<sub>2</sub>O<sub>3</sub> in the spectral region of 400 to 4000 cm<sup>-1</sup>. The characteristic absorption peaks of oxide groups belong to 1388 cm<sup>-1</sup>, 1625 cm<sup>-1</sup>. For the OH<sup>-</sup> group of NiO, peak positions are 2388 cm<sup>-1</sup>. The bending mode of H<sub>2</sub>O bond at 1625 cm<sup>-1</sup> as well as its stretching mode in the range of 3000-3700 cm<sup>-1</sup>, which is almost overlapped by the O-H vibration mode of NiO, CoO and Fe<sub>2</sub>O<sub>3</sub> at 2358 cm<sup>-1</sup>, are observed in the FT-IR analysis of the Oxide materials [9] (Table 3).

### Magnetic studies

The experimental results for NiO, CoO and Fe<sub>2</sub>O<sub>3</sub> are more complete and will, therefore, be treated first. Thus, we note first that the magnetization curves for these samples for the present particle size range (i.e. below about 50 nm) and manifest a peak at about 29

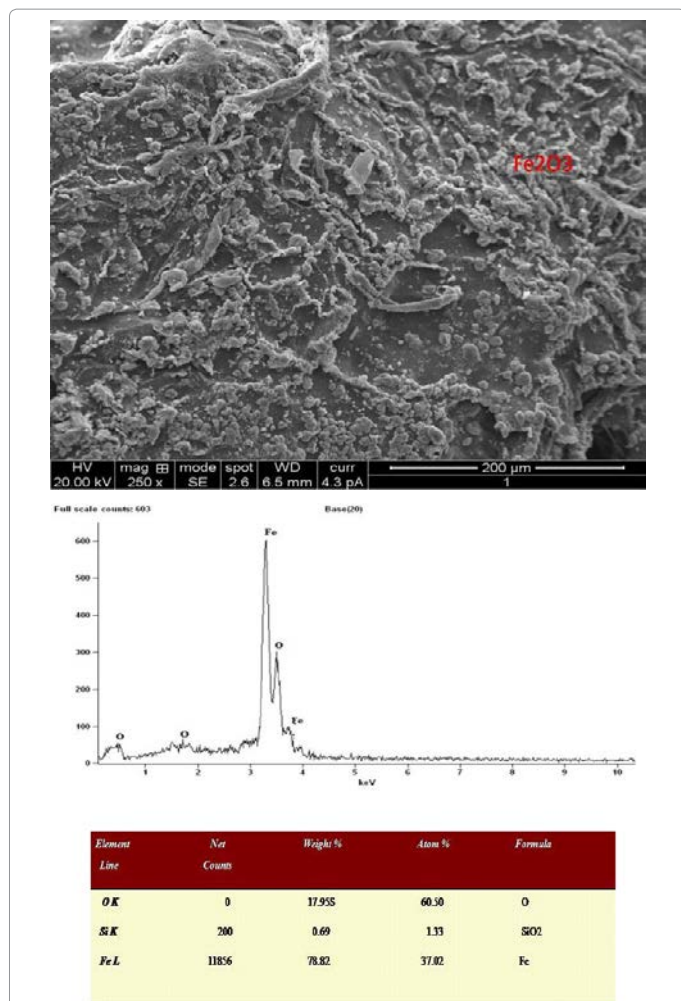


Figure 4: (a and b) EM images and EDAX analysis of Fe<sub>2</sub>O<sub>3</sub> NPs at 250°C.

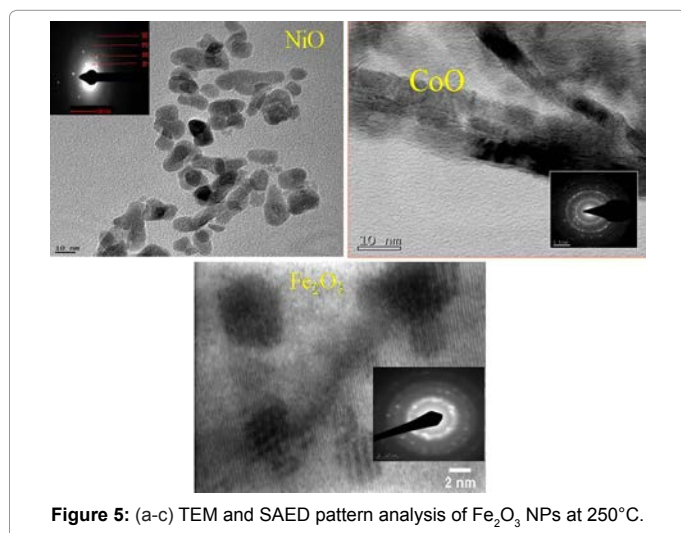


Figure 5: (a-c) TEM and SAED pattern analysis of Fe<sub>2</sub>O<sub>3</sub> NPs at 250°C.

K (which should approximate the Néel temperature, known to be a degree or two smaller than the peak temperature and converge at about 100 K (Table 4). The inverse susceptibility shown in Table 4 manifest an anti-ferromagnetic behavior, namely,  $\chi_{(T)} = \chi_0 + C/(T-\theta)$  with a Weiss

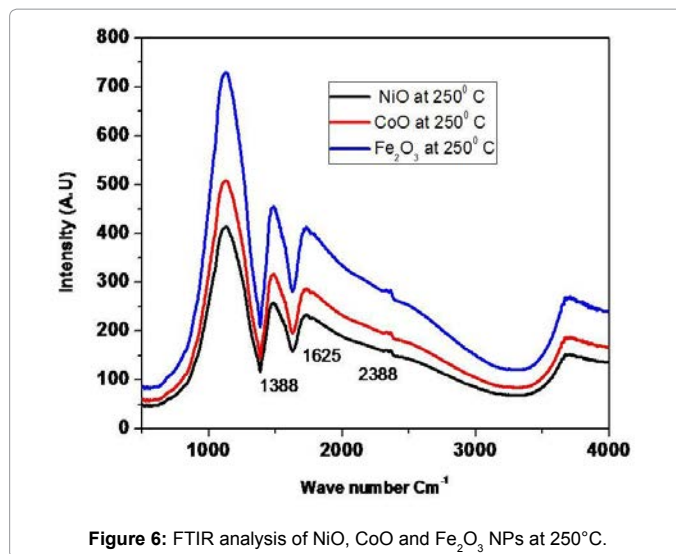


Figure 6: FTIR analysis of NiO, CoO and Fe<sub>2</sub>O<sub>3</sub> NPs at 250°C.

	NiO	CoO	Fe <sub>2</sub> O <sub>3</sub>
Coercivity (H <sub>ci</sub> ) (G)	23.866	25.169	27.160
Magnetization (M <sub>s</sub> ) (emu/g)	52.505	54.826	61.369
Mass (mg)	1.0000	1.0000	1.0000
Retentivity (M <sub>r</sub> ) (×10 <sup>-3</sup> emu/g)	46.173	47.236	48.325

Table 3: Magnetic calculated parameters of NiO, CoO and Fe<sub>2</sub>O<sub>3</sub> nanoparticles annealed at 250°C.

temperature  $\theta < 0$ . Here C is the Curie constant and  $\chi_0$  comprises an admixture of several contributions, including various magnetic phases contributions a temperature independent orbital contribution and a small diamagnetic contribution measured in this system elsewhere to be approximately  $2.06 \times 10^{-6}$  emu/Oe.g for NiO (i.e. relatively small),  $2.53 \times 10^{-6}$  emu/Oe.g, for CoO and  $2.96 \times 10^{-6}$  emu/Oe.g for Fe<sub>2</sub>O<sub>3</sub>. Choosing to find the values of Curie constant for the sample Fe<sub>2</sub>O<sub>3</sub> neglecting  $\chi_0$  yields a Curie constant  $CB \approx 6.54 \times 10^{-3}$  K emu/(g Oe), and hence an effective moment per ion  $\mu_{\text{eff}} \approx 3.69 \mu_B$ , with  $\theta_B \approx -55$  K. The determination of  $\theta$  is, however, known to be problematical and a large range of values is reported in the literature (Figure 7). On the assumption that the overall magnetic behavior  $m(H_0)$  (Figure 7) of this nano-particulate system can be expressed as a linear combination of an average antiferromagnetic behavior for the core component and an average superparamagnetic behavior for the shell component it would be given also Chen et al. [10].

### Antibacterial activity of compounds against bacterial pathogens using well diffusion method

The antibacterial activities were evaluated against *E. coli*, a Bacillus and Pseudomonas. The bacteria were thawed on ice for 30 minutes before being placed on an agar plate. The plate was then dried before incubation for 17 h in a standard cell culture environment (37°C, 7% CO<sub>2</sub>, and 97% air). A single colony of *E. coli* was selected using a 10- $\mu$ L loop and inoculated into a centrifuge tube containing 5 mM of cryptic soy broth. Bacteria in the centrifuge tube were then incubated at 37°C under agitation at 200 rpm for another 15 h. At that point, the bacteria solution was diluted in cryptic soy broth to an optical density of 0.52 at 600 nm using a microplate reader. According to the standard curve correlating bacteria number with optical density, this value was equivalent to  $5 \times 10^6$  cells/mL. The cells were further diluted in cryptic soy broth to  $5 \times 10^4$  cells/mL before being added to a new centrifuge tube at 5 mL/tubes. Concentrated of metal oxides of these three in solution



Microbes	Zone of Inhibition in mm NiO			Zone of Inhibition in mm CoO			Zone of Inhibition in mm Fe <sub>2</sub> O <sub>3</sub>		
	25 µl	50 µl	100 µl	25 µl	50 µl	100 µl	25 µl	50 µl	100 µl
<i>E. coli</i>	0.8	1.6	1.3	0.8	1.4	1.1	0.7	1.0	1.3
<i>Bacillus</i>	1.1	1.0	1.0	0.9	1.1	1.0	0.8	0.8	0.9
<i>Pseudomonas</i>	0.7	0.9	1.1	0.9	1.0	1.1	0.5	0.5	0.6

Table 4: Antibacterial activity analysis details.

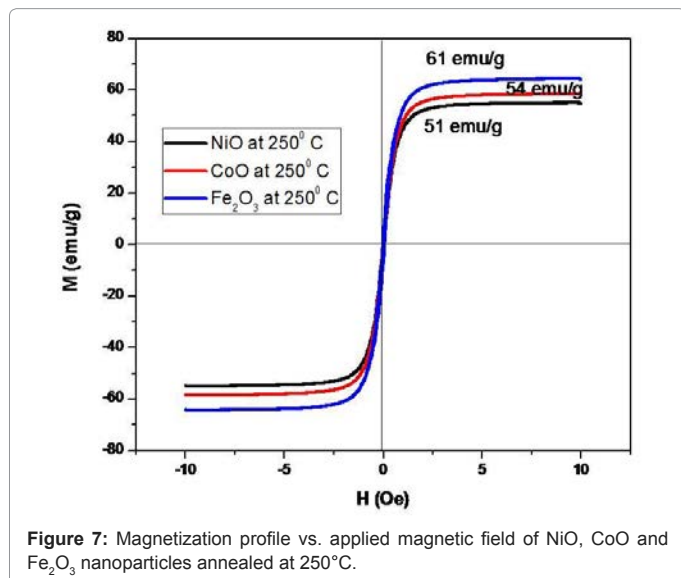


Figure 7: Magnetization profile vs. applied magnetic field of NiO, CoO and Fe<sub>2</sub>O<sub>3</sub> nanoparticles annealed at 250°C.

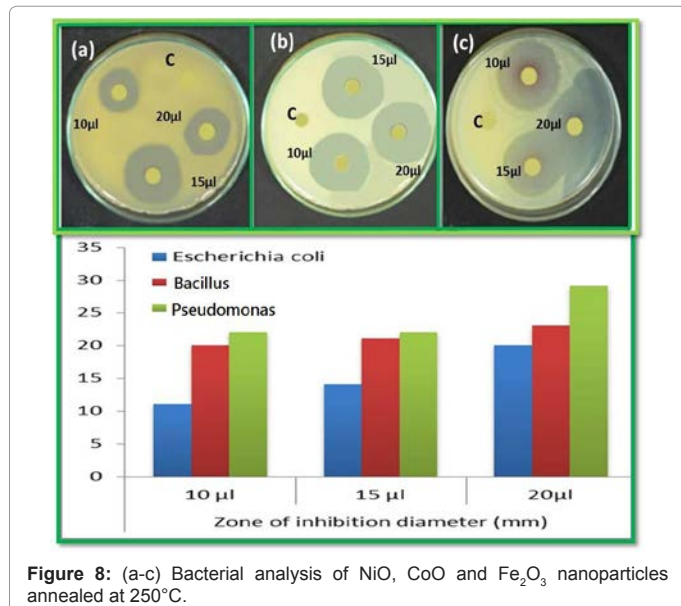


Figure 8: (a-c) Bacterial analysis of NiO, CoO and Fe<sub>2</sub>O<sub>3</sub> nanoparticles annealed at 250°C.

were added to bacteria tubes at different doses (2 mM (low dose), 5 mM (medium dose), and 10 mM (high dose)). A tube of bacteria without nanoparticles served as a control. Bacteria were then incubated under agitation for different concentration solution was transferred to a 100-well plate for optical density readings at 600 nm using a microplate reader. The plates were incubated at 30°C for 24 h, and then the diameters of the zone of inhibition were measured in millimetres. Each antibacterial assay was performed in triplicate and mean values were reported (Figure 8). NiO, Fe<sub>2</sub>O<sub>3</sub> and CoO were treated for antibacterial activity against *E. coli*, *Bacillus* and *Pseudomonas* as shown in the

Figure 8a-8c. These species were grown in Luria Bertaniborth (LB) and Tryptic soy Borth (TSB) respectively and incubated at 35°C for all the assays overnight cultures were used in 1:100 dilutions in Muller Hinton agar (MHA) medium. Wells with a diameter of 3 mm were then punched out of the swabbed plates. Dimethyl sulfoxide (DMSO) was served as control. Then, the plates were incubated at 35°C for 36 h. In the zone of inhibition was measured and the results were expressed in millimetre diameter. NiO, Fe<sub>2</sub>O<sub>3</sub> and CoO, which is attributed to the medium. The antibacterial results showed that for NiO, Fe<sub>2</sub>O<sub>3</sub> and CoO nanocomposite material has some significant effect on the bacterial growth for NiO, Fe<sub>2</sub>O<sub>3</sub> and CoO nanocomposites portrayed an excellent activity against *S. aureus*, *E. hermannii*, then the *P. vulgaris* reverse case was obtained with 1:100 ratios. In the presence of reactive oxygen species (ROS) generated by for NiO, Fe<sub>2</sub>O<sub>3</sub> and CoO nanoparticles are responsible for their bacterial activity. It was further showed that the antibacterial behaviour of for NiO, Fe<sub>2</sub>O<sub>3</sub> and CoO nanocomposites could be due to the chemical interactions between hydrogen and membrane proteins, or other chemical species produced in the presence of for NiO, Fe<sub>2</sub>O<sub>3</sub> and CoO nanoparticles. It was shown in the study that nano sized for NiO, Fe<sub>2</sub>O<sub>3</sub> and CoO nanoparticles are responsible for inhibiting bacterial growth. The photographs of the antimicrobial activity of the NiO, CoO and Fe<sub>2</sub>O<sub>3</sub> composites nanoparticles were investigated against *S. aureus* (MTCC 1430) followed by *E. coli* (MTCC 739) by agar diffusion assay and Resazurin Microtitre Assay (REMA) (Sotirious Baskoutas) [11]. Since *Staphylococcus aureus* and the *Escherichia coli* have been implies to be the important common pathogens in biomaterial-associated infections. However, there are some differences between gram positive *S. aureus* and gram negative *E. coli*. *S. aureus* has slightly less bacterial activities in growth curves, protein leakage, and inactivation of LDH (Lactate Dehydrogenase activity) than *E. coli*. The morphological destruction of bacterial cell of *S. aureus* was weak than *E. coli*. This difference was possibly attributable to the difference of the peptidoglycan layer of the bacterial cell between gram positive *S. aureus* and *E. coli*, an essential function of the peptidoglycan layer is to protect against antibacterial agents such as antibiotics, toxins, chemicals, and cell membrane [12-14]. The oxide materials with different samples of NiO, CoO and Fe<sub>2</sub>O<sub>3</sub> are important factor for antimicrobial activity. Initially NiO sample, CoO sample and Fe<sub>2</sub>O<sub>3</sub> sample, it can be used as an antimicrobial material or coatings in various day to day applications (bath room tiles, doorknobs, packing materials etc.) by controlling the spread of disease causing microbes. Since metal oxides have the good adsorption property and they are well known for its photo-degradation nature the composites can be used as absorbent and degradation agent for the pollutants. Hence the combination for the NiO, CoO and Fe<sub>2</sub>O<sub>3</sub> has excellent applications in biomedical, antimicrobial and environmental friendly applications [15,16]. In case NiO highest dipping rate of sample clearly implies that the bacterial growth decreases and exponentially with increasing dipping rate increases the process up to certain concentration. In such a case, basically *E. coli* has a relatively thin cell wall made of peptidoglycans and lipopolysaccharide [17,18]. It is concluded that increasing the dipping rate due to increase the metal oxides change of inhibited area of zone also increases due to change antimicrobial activity of against *Bacillus* (MTCC 1430) followed by

*E. coli* (MTCC 739) by agar diffusion assay and Resazurin Microtitre Assay (REMA). The initial concentration of the microorganisms used was  $1 \times 10^8$  cells/ml (test culture). The agar diffusion assay is performed on Mueller-Hinter (MH) agar. 1% test culture is seeded into the medium before pouring it in to Sterile Petri plates to form a layer of 4 mm thickness. Metal oxides are loaded on sterile discs (Himedia). Which are placed on the MH agar plates and incubated for 24 h at 37°C. The results are recorded by observing the zone of inhibition as shown in Table 4 [19]. It is good agreement with reported that *S. aureus* has exhibited the most susceptibility to the composites in comparison with *E. coli* [20]. The addition of the outer membrane of the gram-negative bacteria cells influences the permeability of many molecules. Under certain conditions, the gram-negative bacteria are more resistant to many chemical agents than gram-positive cells [21]. The antibacterial activity of the Bacillus spp composites on *E. coli*. The samples were tested for antimicrobial activity by Disc Diffusion Method against pathogenic organisms such as *E. coli* and Bacillus spp. The pure cultures of organisms were sub-cultured on nutrient broth at 35°C on a rotary shaker at 300 rpm. Loop full isolated subculture swabbed on MHA agar plate. The pre culture plate was prepared by standard method [22-24]. After solidification of the agar plate, different types of test pathogens were swabbed in each of the agar plate using sterile cotton buds and labeled clearly. The plates were then supplemented with different composite thin films and incubated at 37°C for 24 h of incubation to observe the zone of inhibition.

## Conclusion

NiO, Fe<sub>2</sub>O<sub>3</sub> and CoO were successfully synthesised by biosynthesis method. The effect of annealing temperatures at 250°C on their comparative structural and morphological properties of metal oxides was investigated using X-ray diffraction and Scanning Electron Microscopy and Transmission Electron Microscopy respectively. In addition, the characterizations, analysis and simulation studies recommended these three unstable metal oxides were very much helpful for magnetic and biological applications.

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